

# Effects of postharvest treatments based on calcium and silicon in hydro-cooling on the basic quality attributes of 'Bing' sweet cherries (*Prunus avium* L.) during storage

## Tratamientos poscosecha a base de calcio y silicio en hidro-enfriamiento sobre atributos básicos de calidad en cerezas (*Prunus avium* L.) dulces 'Bing' durante almacenamiento

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### ABSTRACT

Ca<sup>2+</sup> and Si<sup>2+</sup> treatments confer resistance to biotic and abiotic stresses in many fruits. In sweet cherries, Ca<sup>2+</sup> improves shelf life extension during storage, but only CaCl<sub>2</sub> is used. On the other hand, there is scarce information on CaCO<sub>3</sub> as a source of Ca<sup>2+</sup>, which has shown increased firmness in berries. This study evaluated different treatments based on Ca<sup>2+</sup> (CaCl<sub>2</sub> and CaCO<sub>3</sub>) + Si<sup>2+</sup> (SiO<sub>2</sub>) alone and combined with immersion in hydro-cooling (0°C) on physicochemical characteristics of 'Bing' sweet cherries (*Prunus avium* L.) during storage at low temperature (4°C). Results demonstrate that alone or combined treatments (Ca<sup>2+</sup> and Si<sup>2+</sup>) with hydro-cooling significantly affected skin and flesh color of sweet cherries. Chromaticity (C\*) was increased in treated fruits, indicating an intense red color, especially in those cherries treated with CaCl<sub>2</sub>. Furthermore, firmness was increased during storage in treatments with Ca<sup>2+</sup>, while SiO<sub>2</sub> treatment increased total soluble solids (TSS). Therefore, combined treatments of Ca<sup>2+</sup> and Si<sup>2+</sup> with hydro-cooling might be a promising postharvest strategy to maintain desirable physicochemical characteristics in sweet cherries during low-temperature storage.

### Keywords

*Prunus avium* • fruit firmness • shelf life • non-climacteric fruit • total soluble solids • skin color

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## RESUMEN

Se ha demostrado que los tratamientos con  $\text{Ca}^{2+}$  y  $\text{Si}^{2+}$  confieren resistencia al estrés biótico y abiótico en muchas frutas. En cerezas dulces, el  $\text{Ca}^{2+}$  mejora la extensión de la vida útil durante el almacenamiento, pero solo se ha utilizado  $\text{CaCl}_2$ . Por otro lado, existe escasa información sobre el  $\text{CaCO}_3$  como fuente de  $\text{Ca}^{2+}$ , que ha mostrado un aumento de la firmeza en bayas. En este estudio, se evaluaron diferentes tratamientos a base de  $\text{Ca}^{2+}$  ( $\text{CaCl}_2$  y  $\text{CaCO}_3$ ) +  $\text{Si}^{2+}$  ( $\text{SiO}_2$ ) solos y combinados por inmersión en hidro-enfriamiento ( $0^\circ\text{C}$ ) sobre características fisicoquímicas en cerezas dulces 'Bing' (*Prunus avium* L.) durante el almacenamiento a baja temperatura ( $4^\circ\text{C}$ ). Los resultados demuestran que los tratamientos solos o combinados ( $\text{Ca}^{2+}$  y  $\text{Si}^{2+}$ ) en hidro-enfriamiento afectaron significativamente al color de la piel y pulpa de las cerezas dulces. Se aumentó la cromaticidad ( $C^*$ ) en los frutos tratados, indicando un color rojo intenso, especialmente en aquellas cerezas tratadas con  $\text{CaCl}_2$ . Además, la firmeza aumentó durante el almacenamiento en los tratamientos con  $\text{Ca}^{2+}$ , mientras que el tratamiento con  $\text{SiO}_2$  incrementó la acumulación de sólidos solubles totales (SST). Por lo tanto, los tratamientos combinados de  $\text{Ca}^{2+}$  y  $\text{Si}^{2+}$  con hidro-enfriamiento podrían ser una estrategia poscosecha prometedora para mantener las características fisicoquímicas deseables en cerezas dulces durante el almacenamiento a baja temperatura.

### Palabras clave

*Prunus avium* • firmeza del fruto • vida útil • fruto no climatérico • sólidos solubles totales • color de la piel

## INTRODUCTION

Sweet cherry (*Prunus avium* L.) is one of the most appreciated fruits worldwide. Attributes such as sweetness, color, size, and flavor add up to being a rich source of antioxidants and phytonutrients (14, 39, 40, 66). In Mexico, the current demand for sweet cherries exceeds the 1,249 tons imported (17). In this country, cherry production is 144.45 tons, with only 35.5 ha established in the states of Chihuahua and Puebla (50). However, Mexico has regions with high potential for its production (4).

Fruit firmness, skin and pedicel color, acidity, and sugar content in fresh sweet cherries are major attributes influencing consumer acceptability (14). However, these attributes are often lost in between harvest, packaging, transportation, and storage, especially since sweet cherries are highly perishable and have a shorter post-harvest shelf life (40, 42, 49). Post-harvest strategies should avoid water loss, softening, color deterioration, and pedicel browning (14, 30, 53, 66). Nowadays, several technologies and practices, aimed at preserving post-harvest quality of sweet cherries, target respiration and senescence, increasing flesh firmness (10, 14, 54, 58, 66). In this regard, pre-harvest or at-harvest treatments with calcium ( $\text{Ca}^{2+}$ ) and silicon ( $\text{Si}^{2+}$ ) on sweet cherries extend storage life and improve flesh firmness by minimizing respiration and increasing fruit flesh resistance (14, 16, 31, 33, 46, 58, 63, 64).

Calcium is considered a critical, quality-defining nutrient in sweet cherries (63), mainly promoting firmness by acting in association with pectin molecules at cell-wall level (8, 38).  $\text{CaCl}_2$  is the most widely used source of calcium in sweet cherries, both pre and post-harvest, preserving fruit quality and reducing physiological disorders like cracking (12, 14, 16, 27, 64).  $\text{CaCO}_3$  is another less-known source of calcium for agriculture, shown to increase firmness of 'Shiraz' grapes after pre-harvest foliar application (32). On the other hand, silicon ( $\text{Si}^{2+}$ ), although not considered an essential element for plant nutrition (7), has been suggested against various biotic and abiotic stresses in sweet cherry cultivation (2, 7, 28, 46).  $\text{Si}^{2+}$  improves strength and stiffness of plant tissues and increases wall extensibility (2, 23, 28). In addition, available literature demonstrates the safe use of physical treatments like hydro-cooling on vegetables and fruits to extend postharvest quality, especially by delaying firmness loss, reducing respiration rate and preserving fruit flavor (58, 60).

Therefore, chemical strategies like  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  applications and physical treatments like hydro-cooling on freshly harvested sweet cherries might maintain storage quality (58, 59). However, studies considering a combination of  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  with hydro-cooling and cool storage on post-harvest quality and shelf life of sweet cherries, are scarce (29, 53, 58).

Considering the aforementioned, the study aimed to evaluate the effect of post-harvest treatments with  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  combined with hydro-cooling on physicochemical quality of 'Bing' sweet cherries during low-temperature storage.

## **MATERIALS AND METHODS**

### **Fruits and chemical inputs**

Sweet cherries 'Bing' (12 kg) were harvested from the commercial orchard "El Fulano" (28°26'46" N; 106°45'1.6" W and 2013 m above sea level) located in the "Tres Lagunas" ejido, in Cuauhtemoc, Chihuahua, Mex. Fruits were randomly collected from several trees on east-facing branches and from the center of the canopy. For the treatments of  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$ ; food-grade  $\text{CaCl}_2$ ,  $\text{CaCO}_3$ , and  $\text{SiO}_2$  were purchased from Food Technologies Trading S.A. de C.V. Mexico.

### **Immersion of fruits**

Before starting treatments, cherries were disinfected by immersion in a 1% (v/v) sodium hypochlorite for 5 min, washed twice with sterile distilled water, and left to dry at room temperature while packaged in commercial polyethylene boxes. Six treatments (solutions) simulated hydro-cooling, using distilled water and enough ice to keep the solutions at 0°C (58). Sweet cherries were immersed for 5 min in the evaluated solutions, all of them at 0.5% according to previous studies (58). The evaluated solutions were T1 ( $\text{CaCl}_2$ ), T2 ( $\text{CaCO}_3$ ), T3 ( $\text{SiO}_2$ ), T4 ( $\text{CaCl}_2 + \text{SiO}_2$ ), T5 ( $\text{CaCO}_3 + \text{SiO}_2$ ) and a control treatment T6 (distilled water at 0°C)]. Thirty-two selected fruits were used in each treatment considering post-harvest evaluation dates 0, 7, 14 and 21 days after treatment. After the treatments, fruits were drained, placed on brown paper to dry at room temperature, packed in commercial polyethylene boxes (500 g) and immediately stored at 4°C with relative humidity of ~85%.

### **Basic physicochemical properties**

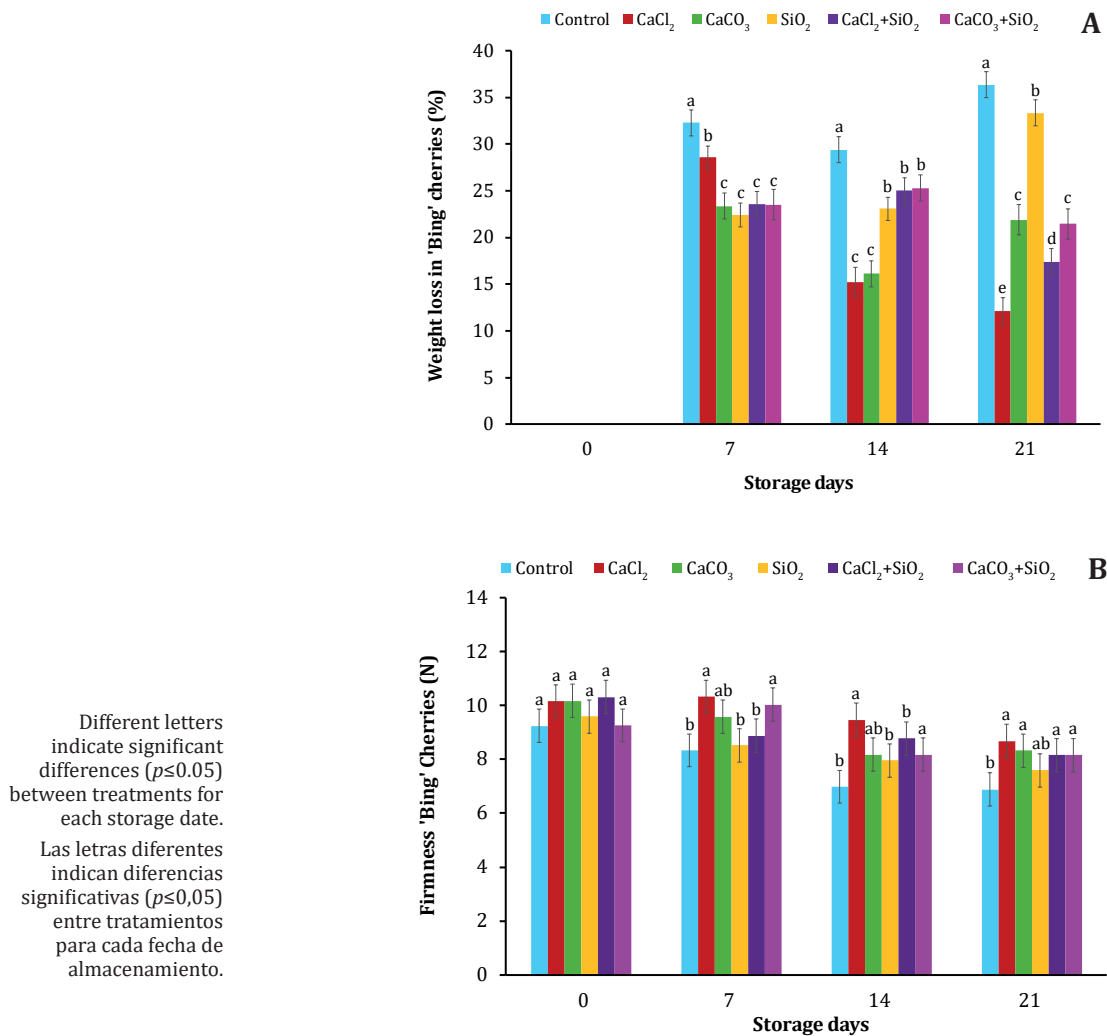
Physicochemical changes were measured by monitoring weight, firmness, color, total soluble solids (TSS; °Brix), and titratable acidity (TA). Measurements were expressed as the average of 32 fruits. The standard error (SE) was estimated at each evaluation time. Fruit weight was determined with an electronic balance, 0.01g precision, Precisa BJ 610C (Precisa Gravimetrics AG/Switzerland). Fruit firmness was evaluated as fruit resistance to a deformation of 15% of fruit diameter using a plunger of  $\varnothing=6$  mm on a stationary steel plate, attached to a Universal Texture Analyzer TA-XT2i (Texture Technologies Corp. USA) according to previous studies (6). Data were expressed in Newtons (N) using the Texture Exponent Lite program. Skin color (CIELab parameters  $L^*$ ,  $C^*$  and  $h^*$ ) was measured at opposite sites of each fruit with a colorimeter CR-300, Minolta, (Japan). Total soluble solids content (TSS= °Brix) was determined in fruit juice with a digital refractometer PAL-1 pocket (Atago, Japan). Finally, titratable acidity (TA expressed as g 100 g<sup>-1</sup> of fresh weight 'FW') was measured by diluting 1 g of flesh in 9 mL of distilled water, followed by 3 drops of phenolphthalein and titrated with 0.1 N NaOH until pH 8.2 (6). The maturity index was expressed as the ratio of TSS: TA (34).

### **Experimental design and statistical analysis**

Results were statistically evaluated according to a split-plot in-time design. ANOVA and LSD mean tests were used to detect significant differences among treatments at  $p \leq 0.05$  using SAS System for Windows 9.0 (SAS Institute. Inc. Cary, N.C., USA, 2002) after testing assumptions. All experiments were conducted using four replicates.

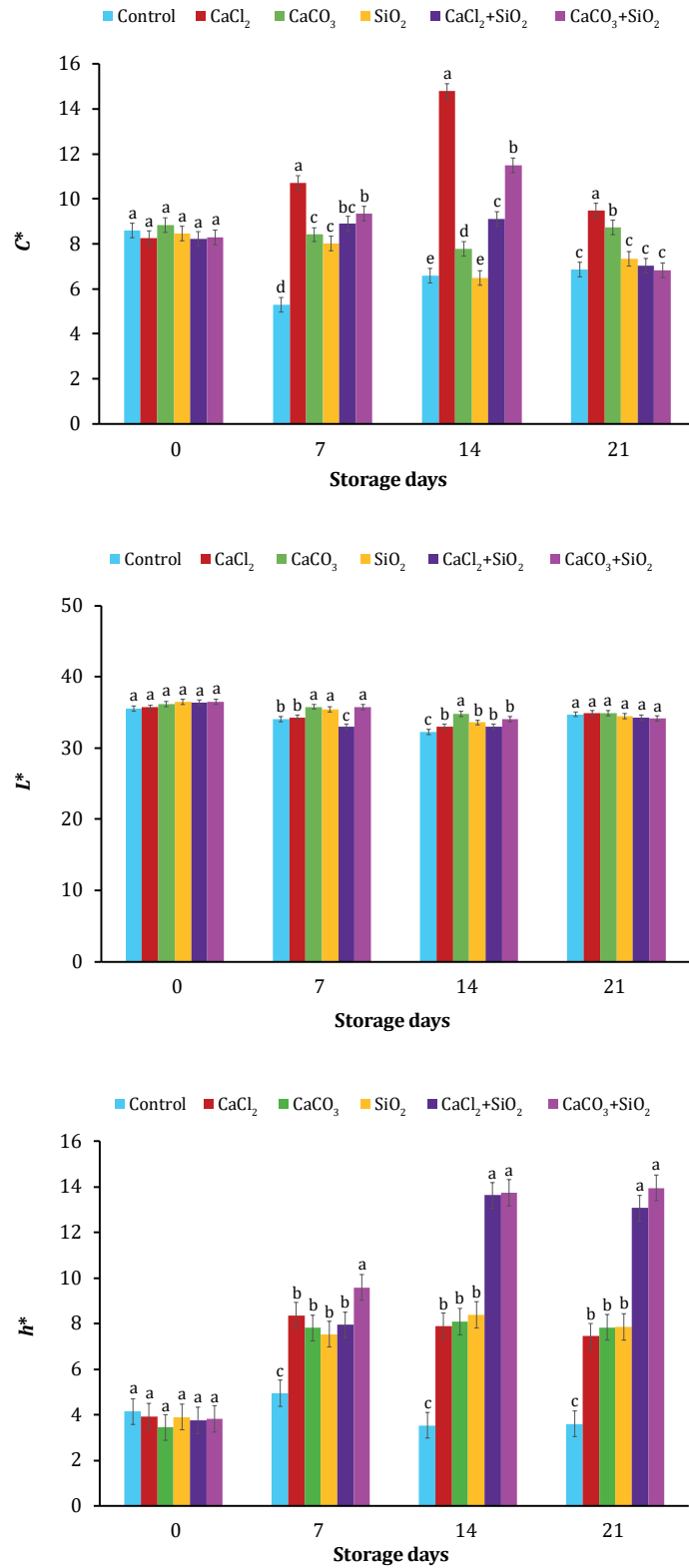
RESULTS AND DISCUSSION

In hydro-cooling, calcium and silicon treatments (alone or combined) significantly influenced some quality parameters and shelf life in sweet cherries during low-temperature storage (figure 1, figure 2, page XXX and figure 3, page XXX). Various studies have extensively documented that  $Ca^{2+}$  applications in fruits favor storage conservation. In sweet cherries, it has been documented that  $Ca^{2+}$  delays deterioration, favorably influencing physicochemical attributes like weight, color, firmness, TSS, TA, pH, respiration rate, and anthocyanin content, especially during storage (14, 31, 57, 58, 59, 60). Shelf life extension in sweet cherries could be attributed to  $Ca^{2+}$  increase in the cell walls, favored by rapid absorption of  $Ca^{2+}$  by the fruit flesh under hydro-cooling immersion (19, 27, 59, 61).



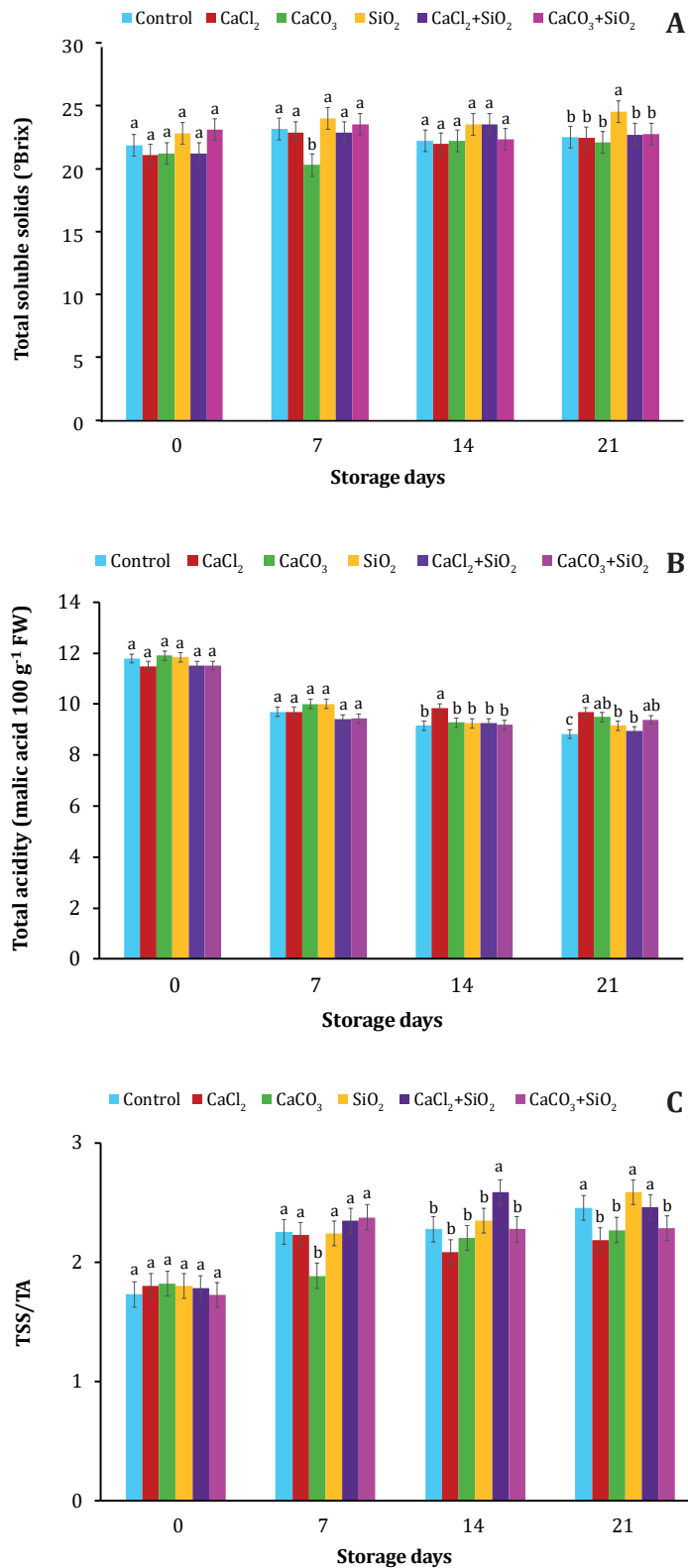
**Figure 1.** Effect of post-harvest treatments based on calcium ( $Ca^{2+}$ ) and silicon ( $Si^{2+}$ ) sources, alone and combined with hydro-cooling on weight loss (A) and firmness (B) in 'Bing' sweet cherries during storage at low temperature.

**Figura 1.** Efecto de los tratamientos poscosecha basados en fuentes de calcio ( $Ca^{2+}$ ) y silicio ( $Si^{2+}$ ) solas y combinadas con hidro-enfriamiento sobre la pérdida del peso (A) y la firmeza (B) en cerezas dulces 'Bing' durante el almacenamiento a baja temperatura.



**Figure 2.** Effect of post-harvest treatments of calcium ( $\text{Ca}^{2+}$ ) and silicon ( $\text{Si}^{2+}$ ) sources alone and/or combined with hydro-cooling on skin color ( $L^* C^* h^\circ$ ) in 'Bing' sweet cherries during low-temperature storage.

**Figura 2.** Efecto de los tratamientos poscosecha de fuentes de calcio ( $\text{Ca}^{2+}$ ) y silicio ( $\text{Si}^{2+}$ ) solas y/o combinadas con hidro-enfriamiento sobre el color de la piel ( $L^* C^* h^\circ$ ) en cerezas 'Bing' dulces durante el almacenamiento a baja temperatura.



**Figure 3.** Effect of post-harvest treatments of calcium ( $\text{Ca}^{2+}$ ) and silicon ( $\text{Si}^{2+}$ ) sources alone and/or combined with hydro-cooling on total soluble solids (TSS; A), titratable acidity (TA; B) and maturity index (TSS/ TA; C) in 'Bing' sweet cherries during low-temperature storage.

**Figura 3.** Efecto de los tratamientos poscosecha de fuentes de calcio ( $\text{Ca}^{2+}$ ) y silicio ( $\text{Si}^{2+}$ ) solas y/o combinadas con hidro-enfriamiento sobre los sólidos solubles totales (SST; A), la acidez titulable (AT; B) y el índice de madurez (SST/AT; C) en cerezas dulces 'Bing' durante el almacenamiento a baja temperatura.

Weight loss is the most important parameter for horticultural crops and fruit quality and shelf life. All treatments based on  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  sources, alone and combined with hydro-cooling, affected weight loss of sweet cherries during storage (figure 1, page XXX). According to previous studies (51, 66), weight loss in stored fruits mainly depends on transpiration and respiration. Interestingly, cherries treated with  $\text{Ca}^{2+}$  lost less weight during storage compared to untreated cherries (figure 1, page XXX), suggesting that  $\text{Ca}^{2+}$  ions increased cell wall stability. Other studies mention increased cell wall stability after  $\text{Ca}^{2+}$  ions bind non-esterified pectins and stabilize cell membranes, preventing electrolyte leakage and consequently preventing fruit moisture and weight loss (1, 38, 41). The observed weight values in fruits treated with  $\text{Ca}^{2+}$  could have been influenced by the amount of this element absorbed through the skin (through the lenticels and peduncle pores) during the 5-minutes exposure (44). Similarly, previous studies (15) documented that combined Ca-Glu (calcium gluconate) treatment, limited weight loss in sweet cherries.

Sweet cherries treated with  $\text{SiO}_2$  showed rapid weight loss on day 21 of storage, however less evident than for control fruits (figure 1A, page XXX). Similarly, other studies (3) have documented that  $\text{SiO}_2$  was less effective in preventing weight loss in post-harvest fruits of *Citrus × sinensis*, while Rombolà *et al.* (2023) found that foliar sprays with sodium silicate ( $\text{Na}_2\text{SiO}_3$ ) decreased cherry weight at harvest.

Firmness is a major attribute in fruits (43). Broadly, our study showed a gradual loss of firmness concerning storage time indicating senescence, with significant differences among monitoring dates and treatments. According to previous studies (14), decreases in this parameter are more noticeable during storage. Softening of sweet cherries is attributed to enzymatic degradation of pectic compounds in the middle lamella of the cell walls by polygalacturonases, pectin methyl esterases, cellulases, and  $\beta$ -galactosidases (62). All sweet cherries treated with  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  were firmer than control fruits (figure 1B, page XXX). Studies have suggested that pre- and post-harvest treatments with  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  favor greater firmness in fruits at harvest time and during storage (27, 55). Sweet cherries containing insufficient  $\text{Ca}^{2+}$  are softer, and, therefore, more susceptible to quality losses during storage (10). Fruits treated with  $\text{CaCl}_2$  were the firmest compared to control fruits after 21 days of storage (figure 1B, page XXX). It has been evidenced that  $\text{CaCl}_2$  applied before and/or after cherry harvest increases firmness values up to 0.6 N (14, 63, 64). Our study is consistent with previous studies (10, 14, 15, 27, 55), reporting increased fruit firmness in treatments with  $\text{Ca}^{2+}$  before harvest and/or in recently harvested cherries. The treatments ( $\text{CaCO}_3$  and  $\text{CaCO}_3+\text{SiO}_2$ ) also favored greater firmness of sweet cherries but to a lesser extent than  $\text{CaCl}_2$  (figure 1B, page XXX). Similarly, other studies (32) documented firmer 'Shiraz' grapes after pre-harvest foliar treatment with  $\text{CaCO}_3$ . In our study, the treatment with  $\text{SiO}_2$  alone was the least effective, although slightly superior to the control.

The greater firmness of sweet cherries treated with  $\text{Ca}^{2+}$  is attributed to the ability of this element to maintain cell wall mechanical properties and integrity during storage, which consequently delays softening (14, 44, 47). According to previous studies (38),  $\text{Ca}^{2+}$  acts in association with pectin molecules in fruit cell walls. It has also been suggested that  $\text{Ca}^{2+}$  maintains fruit firmness by reducing water loss and stabilizing the membrane, given this ion is responsible for binding phosphate and carboxylate groups of membrane phospholipids and proteins (62, 65).

Surface color of cherries is determined by factors such as radiation at the end of fruit development, and temperatures near harvest (13). Recently, it has been documented that color of sweet cherries is influenced by post-harvest treatments based on  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  (14, 46). On the other hand, according to other studies (21, 36), the chromatic functions  $L^*$ ,  $C^*$  and  $h^\circ$  are closely correlated with color change and anthocyanin accumulation in sweet cherries during ripening. Interestingly, after 21 days of storage, sweet cherries treated with  $\text{Ca}^{2+}$  or  $\text{Si}^{2+}$  showed increased chromaticity (figure 2, page XXX), redder and intensity ( $C^*$ ), especially in cherries treated with  $\text{CaCl}_2$ . This effect could be due to the inhibition of skin color development by  $\text{Ca}^{2+}$  or  $\text{Si}^{2+}$ . The delayed skin color darkening may be related to senescence inhibition (58, 59). Control fruits showed a darker red color attributed to chlorophyll degradation and accumulation of anthocyanins during storage (5, 18). Coincidentally, other studies (21) reported that the higher the anthocyanin content in sweet cherries, the lower the values of  $L^*$  and  $h^\circ$ .



The  $L^*$  value in sweet cherries decreased during storage in all treatments, not showing significant differences among treatments (figure 2, page XXX). Sweet cherries treated with  $\text{CaCO}_3+\text{SiO}_2$  and  $\text{CaCl}_2+\text{SiO}_2$  showed a higher  $h^\circ$  angle (figure 2, page XXX), indicating reduced red tones ( $h^\circ$ ) than control fruits and suggesting lower skin anthocyanin content (21, 37). In contrast, Rombolà *et al.* (2023) documented that  $\text{Si}^{2+}$  reduced hue ( $h^\circ$ ), brightness ( $C$ ), and saturation of cherry skin/flesh, while, Karagiannis *et al.* (2021) documented that foliar sprays with  $\text{Si}^{2+}$  induced skin color development in apples by stimulating anthocyanin accumulation. In this experiment, sweet cherries treated with  $\text{CaCO}_3+\text{SiO}_2$  and  $\text{CaCO}_3$  showed higher  $L^*$  and  $h^\circ$  values (figure 2, page XXX) compared with control fruits, probably given to suppression of respiratory processes by  $\text{CaCO}_3$ , as previously established in cherries treated with  $\text{Ca}^{2+}$  at harvest (14). The positive effect of  $\text{CaCO}_3$  on skin and flesh color in sweet cherries is given by  $\text{Ca}^{2+}$  activation of ABA biosynthesis, which influences anthocyanin biosynthesis in non-climacteric fruits such as cherries (20, 32).

The TSS concentration in sweet cherries significantly increased according to storage time in all treatments (figure 3A, page XXX). Increasing TSS concentrations during storage is only frequent in climacteric fruits (22, 35). Therefore, the highest TSS concentrations in non-climacteric sweet cherries might be favored by a pronounced weight/moisture loss in  $\text{SiO}_2$  treated and control fruits (figure 1A, page XXX). The  $\text{SiO}_2$  and  $\text{CaCl}_2+\text{SiO}_2$  treatments significantly increased TSS in sweet cherries (figure 3A, page XXX), like previously documented by Rombolà *et al.* (2023), who suggested that  $\text{Si}^{2+}$  forms a protective film covering fruit surface and preventing transpiration, slowing down phloem translocation, and subsequent sugar accumulation. The high concentration of TSS (figure 3A, page XXX) in  $\text{SiO}_2$ -treated fruits might also be due to sugar concentration after greater weight loss (figure 1A, page XXX) (11), something not observed in  $\text{CaCl}_2$  treated ones.

On the contrary, lower TSS values were observed in sweet cherries treated with  $\text{CaCl}_2$  compared with control fruits. This coincides with other studies (9, 15), documenting low TSS contents in  $\text{Ca}^{2+}$ -treated cherries. Both studies attributed these results to lower respiration rates in treated cherries, leading to cell wall and membrane stabilization. This could also be attributed to delayed moisture and weight loss (figure 1A, page XXX) after pectin stabilization and consequent effects on cell wall and membrane structure (32).

TA in sweet cherries also decreased over time during storage for control,  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  treatments evidencing significant differences (figure 3B, page XXX). Similar results were documented in 'Sweetheart' and 'Lapins' sweet cherries during storage (58). Low acidity mainly depends on ripeness state (45); however, during storage, organic acids might be used as carbon source during respiration (15, 25, 26, 60). After 21 days of storage, sweet cherries treated with  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  maintained TA above values recorded for control cherries. However, the highest TA values were measured in  $\text{CaCl}_2$ -treated fruits (figure 3B, page XXX). Sweet cherries treated with  $\text{CaCO}_3$  and  $\text{CaCO}_3+\text{SiO}_2$  also showed high TA values. Coincidentally, treatments with  $\text{Ca}^{2+}$  (such as  $\text{CaCl}_2$  and Ca-Glu/calcium gluconate) in pre-harvest and/or before storage of sweet cherries, also preserved or retarded TA loss during storage, compared to control fruits (14, 15, 48, 55, 58).

Delayed loss of TA during storage of sweet cherries treated with  $\text{Ca}^{2+}$  sources could be due to the suppressive effect on fruit metabolic activity, especially respiration (15, 35, 56).

The maturity index TSS/TA indicates commercial and organoleptic maturity of fruits (34, 45). High contents of both TSS and TA are associated with good flavor in sweet cherries (52, 53). The TSS/TA ratios in 'Bing' sweet cherries treated with  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  were statistically different (figure 3C, page XXX), however increasing over time in all treatments and indicating a higher acid vs. sugar content ratio. TSS/TA ratio in sweet cherries treated with  $\text{CaCO}_3+\text{SiO}_2$ ,  $\text{CaCO}_3$ , and  $\text{CaCl}_2$  remained lower than control after 21 days of storage, indicating diminished respiration rates. While TSS/TA ratios in  $\text{SiO}_2$  treatments remained above control values.



## CONCLUSIONS

Immersion of freshly harvested 'Bing' sweet cherries with hydro-cooled solutions of  $\text{Ca}^{2+}$  ( $\text{CaCl}_2$  and  $\text{CaCO}_3$ ) and  $\text{Si}^{2+}$  ( $\text{SiO}_2$ ) alone and combined markedly improved quality properties and extended storage capacity at low temperatures. All treatments based on  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  alone reduced weight loss while maintaining firmness, and acidity in sweet cherries. Skin color of sweet cherries treated with  $\text{Ca}^{2+}$  and  $\text{Si}^{2+}$  was more intense than control fruits. Sweet cherries treated with  $\text{CaCl}_2$  were the firmest and had the highest TA values.  $\text{SiO}_2$  increased TSS concentration in sweet cherries, while  $\text{CaCl}_2$  decreased it.

## REFERENCES

- Angeletti, P.; Castagnasso, H.; Miceli, E.; Terminiello, L.; Concellon, A.; Chaves, A.; Vicente, A. R. 2010. Effect of preharvest calcium applications on postharvest quality, softening, and cell wall degradation of two blueberry (*Vaccinium corymbosum*) varieties. *Postharvest Biology and Technology*. 58(2): 98-103. <https://doi.org/10.1016/j.postharvbio.2010.05.015>
- Bat-Erdene, O.; Szegő, A.; Gyöngyik, M.; Mirmazloum, I.; Papp, I. 2021. Effects of silicon in plants with particular reference to horticultural crops - Review article. *International Journal of Horticultural Science*. 27: 95-105. <https://doi.org/10.31421/ijhs/27/2021/9096>
- Beltrán, R.; Otesinova, L.; Cebrián, N.; Zornoza, C.; Breijo, F.; Reig, J.; Garmendia, A.; Merle, H. 2021. Effect of chitosan and silicon oxide treatments on postharvest Valencia Late (*Citrus × sinensis*) fruits. *Journal of Plant Science and Phytopathology*. 5: 065-071. <https://doi.org/10.29328/journal.jpssp.1001063>
- Chávez-Gutiérrez, N. A.; López de Santana-Pimienta, J. A.; Juárez-Méndez, J. 2023. Zonificación agroecológica del cerezo (*Prunus avium* L.) en la región manzanera del estado de Chihuahua. *Ciencia Latina Revista Científica Multidisciplinar*. 7(2): 8683-8709. [https://doi.org/10.37811/cl\\_rcm.v7i2.5983](https://doi.org/10.37811/cl_rcm.v7i2.5983)
- Cogo, S. F.; Chaves, F.; Schirmer, M.; Zambiasi, R.; Nora, L.; Silva, J.; Rombaldi, C. 2011. Low soil water content during growth contributes to preservation of green colour and bioactive compounds of cold-stored broccoli (*Brassica oleraceae* L.) florets. *Postharvest Biology and Technology*. 60(2): 158-163. <https://doi.org/10.1016/j.postharvbio.2010.12.008>
- Correia, S.; Queirós, F.; Ribeiro, C.; Vilela, A.; Aires, A.; Barros, A. I.; Schouten, R.; Silva, A. P.; Gonçalves, B. 2019. Effects of calcium and growth regulators on sweet cherry (*Prunus avium* L.) quality and sensory attributes at harvest. *Scientia Horticulturae*. 248: 231-240. <https://doi.org/10.1016/j.scienta.2019.01.024>
- Coskun, D.; Deshmukh, R.; Sonah, H.; Menzies, J. G.; Reynolds, O.; Ma, J. F.; Kronzucker, H. J.; Bélanger, R. R. 2019. The controversies of silicon's role in plant biology. *New Phytologist*. 221: 67-85. <https://doi.org/10.1111/nph.15343>
- Daher, F. B.; Braybrook, S. A. 2015. How to let go: Pectin and plant cell adhesion. *Frontiers in Plant Sciences*. 6: 1-8. <https://doi.org/10.3389/fpls.2015.00523>
- Díaz-Mula, H. M.; Valero, D.; Guillén, F.; Valverde, J. M.; Zapata, P.J.; Serrano, M. 2017. Postharvest treatment with calcium delayed ripening and enhanced bioactive compounds and antioxidant activity of 'Cristalina' sweet cherry. *Acta Horticulturae*. 1161: 511-514. <https://doi.org/10.17660/ActaHortic.2017.1161.81>
- Dong, Y.; Zhi, H.; Wang, Y. 2019. Cooperative effects of pre-harvest calcium and gibberellic acid on tissue calcium content, quality attributes, and in relation to postharvest disorders of late maturing sweet cherry. *Scientia Horticulturae*. 246: 123-128. <https://doi.org/10.1016/j.scienta.2018.10.067>
- Dutra de Vargas, A.; de Oliveira, F. L.; Quintão Teixeira, L. J.; Oliveira Cabral, M.; dos Santos Gomes Oliveira, L.; Ferreira Pedrosa, J. L. 2022. Physical and chemical characterization of yacon (*Smallanthus sonchifolius*) roots cultivated with different doses of potassium fertilization. *Revista de la Facultad de Ciencias Agrarias. Universidad Nacional de Cuyo. Mendoza. Argentina*. 54(2): 22-31. DOI: <https://doi.org/10.48162/rev.39.079>
- Ekinci, N.; Özdüven, F.; Gür, E. 2016. Effects of preharvest foliar calcium applications on the storage quality of '0900 Ziraat' sweet cherry cultivar. *Erwerbs-Obstbau*. 58: 227-231.
- Ellena, D. 2012. Formación y sistemas de conducción del cerezo dulce. Temuco: Boletín INIA - Instituto de Investigaciones Agropecuarias. N° 247. <https://hdl.handle.net/20.500.14001/7500> (Accessed: 22 February 2023).
- Erbaş, D.; Koyuncu, M. A. 2022. Effect of preharvest calcium chloride treatment on some quality characteristics and bioactive compounds of sweet cherry cultivars. *Journal of Agricultural Sciences (Tarim Bilimleri Dergisi)*. 28(3): 481-489. <http://doi.org/10.15832/ankutbd.874567>
- Erbaş, D.; Koyuncu, M. A. 2023. The effect of pre- and postharvest calcium gluconate treatments on physicochemical characteristics and bioactive compounds of sweet cherry during cold storage. *Food Science and Technology International*. 29(4): 299-309. <https://doi.org/10.1177/10820132221077515>

16. Erogul, D. 2014. Effect of preharvest calcium treatments on sweet cherry fruit quality. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*. 42(1): 150-153. <https://doi.org/10.15835/nbha4219369>
17. FAOSTAT. 2023. Producción mundial de cereza 2021. <https://www.fao.org/faostat/es/#data/QCL/visualize> (Accessed 13 April 2023).
18. Ferrer, A.; Remón, S.; Negueruela, A.; Oria, R. 2005. Changes during ripening of the very late season Spanish peach cultivar Calanda: Feasibility of using CIELAB coordinates as maturity indices. *Scientia Horticulturae*. 105(4): 435-446. <https://doi.org/10.1016/j.scienta.2005.02.002>
19. Figueroa, C.; Opazo, M. C.; Vera, P.; Arraigada, O.; Díaz, M.; Moya-León, M. 2012. Effect of postharvest treatment of calcium and auxin on cell wall composition and expression of cell wall modifying genes in the Chilean strawberry (*Fragaria chiloensis*) fruit. *Food Chemistry*. 132(4): 2014-2022. <https://doi.org/10.1016/j.foodchem.2011.12.041>
20. Gao, Q.; Xiong, T.; Li, X.; Chen, W.; Zhu, X. 2019. Calcium and calcium sensors in fruit development and ripening. *Scientia Horticulturae*. 253: 412-421. <https://doi.org/10.1016/j.scienta.2019.04.069>
21. Gonçalves, B.; Silva, A. P.; Moutinho-Pereira, J.; Bacelar, E.; Rosa, E.; Meyer, A. S. 2007. Effect of ripeness and postharvest storage on the evolution of colour and anthocyanins in cherries (*Prunus avium* L.). *Food Chemistry*. 103: 976-984. <https://doi.org/10.1016/j.foodchem.2006.08.039>
22. Hernández-Muñoz, P.; Almenar, E.; Ocio, M.; Gavara, R. 2006. Effect of calcium dips and chitosan coatings on postharvest life of strawberries (*Fragaria x ananassa*). *Postharvest Biology and Technology*. 39(3): 247-253. <https://doi.org/10.1016/j.postharvbio.2005.11.006>
23. Hossain, M. T.; Mori, R.; Soga, K.; Wakabayashi, K.; Kamisaka, S.; Fujii, S.; Yamamoto, R.; Hoson, T. 2002. Growth promotion and an increase in cell wall extensibility by silicon in rice and some other Poaceae seedlings. *Journal of Plant Research*. 115: 0023-0027. <https://doi.org/10.1007/s102650200004>
24. Karagiannis, E.; Michailidis, M.; Skodra, C.; Molassiotis, A.; Tanou, G. 2021. Silicon influenced ripening metabolism and improved fruit quality traits in apples. *Plant Physiology and Biochemistry*. 166: 270-277. <https://doi.org/10.1016/j.plaphy.2021.05.037>
25. Kays, S. J.; Paull, R. E. 2004. *Postharvest biology*. Athens, GA: Exon Press. <http://hdl.handle.net/10125/65829>
26. Lancho, O.; Velandia, G.; Fischer, G.; Varela, N.; García, H. 2007. Comportamiento de la uchuva (*Physalis peruviana* L.) en poscosecha bajo condiciones de atmósfera modificada activa. *Ciencia y Tecnología Agropecuaria*. 8(1): 61-68. [https://doi.org/10.21930/rcta.vol8\\_num1\\_art:84](https://doi.org/10.21930/rcta.vol8_num1_art:84)
27. Lidster, P. D.; Porritt, S. W.; Tung, M. A. 1978. Texture modification of 'Van'Sweet cherries by postharvest calcium treatments. *Journal of the American Society for Horticultural Science*. 103(4): 527-530. <https://doi.org/10.21273/JASHS.103.4.527>
28. Ma, J. F.; Yamaji, N. 2006. Silicon uptake and accumulation in higher plants. *Trends in Plant Science*. 11: 392-397. <https://doi.org/10.1016/j.tplants.2006.06.007>
29. Manganaris, G. A.; Ilias, I. F.; Vasilakakis, M.; Mignani, I. 2007. The effect of hydrocooling on ripening related quality attributes and cell wall physicochemical properties of sweet cherry fruit (*Prunus avium* L.). *International Journal of Refrigeration*. 30: 1386-1392. <https://doi.org/10.1016/j.ijrefrig.2007.04.001>
30. Martínez-Romero, D.; Alburquerque, N.; Valverde, J. M.; Guillén, F.; Castillo, S.; Valero, D.; Serrano, M. 2006. Postharvest sweet cherry quality and safety maintenance by *Aloe vera* treatment: a new edible coating. *Postharvest Biology and Technology*. 39: 93-100. <https://doi.org/10.1016/j.postharvbio.2005.09.006>
31. Matteo, M.; Zoffoli, J. P.; Ayala, M. 2022. Calcium sprays and crop load reduction increase fruit quality and postharvest storage in sweet cherry (*Prunus avium* L.). *Agronomy*. 12: 829. <https://doi.org/10.3390/agronomy12040829>
32. Maya-Meraz, I. O.; Ornelas-Paz, J. J.; Pérez-Martínez, J. D.; Gardea-Béjar, A. A.; Rios-Velasco, C.; Ruiz-Cruz, S.; Pérez-Leal, R.; Virgen-Ortiz, J. J. 2023. Foliar application of CaCO<sub>3</sub>-rich industrial residues on 'Shiraz' vines improves the composition of phenolic compounds in grapes and aged wine. *Foods*. 12: 1566. <https://doi.org/10.3390/foods12081566>
33. Mitre, V.; Erzsébet, B. U.; Lukacs, L.; Mitre, I.; Teodorescu, R.; Dorel, H. O.; Sestras, A. F.; Stănică, F. 2018. Management of apple scab and powdery mildew using bicarbonate salts and other alternative organic products with fungicide effect in apple cultivars. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*. 46(1): 115-121. <https://doi.org/10.15835/nbha46110783>
34. Monte Andrade, A. D.; Moura, E. A.; Mendonça, V.; Mendes Oliveira, L.; Souza Ferreira, E.; Ferreira Melo, B. E.; Andrade Figueiredo, F. R.; Ferreira Melo, M.; Freitas Medeiros Mendonça, L. 2022. Production and physicochemical characterization of genotypes of *Eugenia uniflora* L. *Revista de la Facultad de Ciencias Agrarias*. Universidad Nacional de Cuyo. Mendoza, Argentina. 54(2): 1-11. DOI: <https://doi.org/10.48162/rev.39.077>
35. Moradinezhad, F.; Ghesmati, M.; Khayyat, M. 2019. Postharvest calcium salt treatment of fresh jujube fruit and its effects on biochemical characteristics and quality after cold storage. *Journal of Horticultural Research*. 27(2): 39-46. <https://doi.org/10.2478/johr-2019-0009>

36. Mozetič, B.; Simčič, M.; Trebše, P. 2006. Anthocyanins and hydroxycinnamic acids of Lambert Compact cherries (*Prunus avium* L.) after cold storage and 1-methylcyclopropene treatment. *Food Chemistry*. 97(2): 302-309. <https://doi.org/10.1016/j.foodchem.2005.04.018>
37. Opiyo, A. M.; Ying, T. J. 2005. The effects of 1-methylcyclopropene treatment on the shelf life and quality of cherry tomato (*Lycopersicon esculentum* var. *cerasiforme*) fruit. *International Journal of Food Science & Technology*. 40: 665-673. <https://doi.org/10.1111/j.1365-2621.2005.00977.x>
38. Ornelas-Paz, J. J.; Quintana-Gallegos, B. M.; Escalante-Minakata, P.; Reyes-Hernández, J.; Pérez-Martínez, J. D.; Ríos-Velasco, C.; Ruiz-Cruz, S. 2018. Relationship between the firmness of Golden delicious apples and the physicochemical characteristics of the fruits and their pectin during development and ripening. *Journal of Food Science and Technology*. 55: 33-41. <https://doi.org/10.1007/s13197-017-2758-6>
39. Ozturk, B.; Aglar, E.; Karakaya, O.; Saracoglu, O.; Gun, S. 2019. Effects of preharvest GA<sub>3</sub>, CaCl<sub>2</sub> and modified atmosphere packaging treatments on specific phenolic compounds of sweet cherry. *Turkish Journal of Food and Agriculture Sciences*. 1(2): 44-56. <https://doi.org/10.14744/turkjfas.2019.009>
40. Parsa, Z.; Roozbehi, S.; Hosseinifarahi, M.; Radi, M.; Amiri, S. 2021. Integration of pomegranate peel extract (PPE) with calcium sulphate (CaSO<sub>4</sub>): A friendly treatment for extending shelf-life and maintaining postharvest quality of sweet cherry fruit. *Journal of Food Processing and Preservation*. 45:e15089. <https://doi.org/10.1111/jfpp.15089>
41. Pérez, A. R.; Quintero, E. M. 2015. Funciones del calcio en la calidad poscosecha de frutas y hortalizas: Una revisión. *Alimentos hoy*. 23(34): 13-25.
42. Petriccione, M.; De Sanctis, F.; Pasquariello, M. S.; Mastrobuoni, F.; Rega, P.; Scortichini, M.; Mencarelli, F. 2015. The effect of chitosan coating on the quality and nutraceutical traits of sweet cherry during postharvest life. *Food and Bioprocess Technology*. 8: 394-408. <https://doi.org/10.1007/s11947-014-1411-x>
43. Pugliese, M. B.; Guzmán, Y.; Pacheco, D.; Bottini, R.; Travaglia, C.; Avenant, J. H.; Avenant, E.; Berli, F. 2022. Indole-3-butyric acid, an alternative to GA<sub>3</sub> for bunch quality enhancing of table grape *Vitis vinifera* L. cv. Superior Seedless. *Revista de la Facultad de Ciencias Agrarias. Universidad Nacional de Cuyo. Mendoza. Argentina*. 54(1): 163-174. DOI: <https://doi.org/10.48162/rev.39.075>
44. Reyes-Medina, A. J.; Pinzon, E. H.; Alvarez-Herrera, J. G. 2017. Effect of calcium chloride and refrigeration on the quality and organoleptic characteristics of cape gooseberry (*Physalis peruviana* L.). *Acta Agronómica*. 66(1): 15-20. <https://doi.org/10.15446/acag.v66n1.50610>
45. Rodrigues Gomes, F.; Morais Silveira, C.; Marques Rodrigues, C. D.; Alves Ferreira, B.; Lopes Barros, Â.; Hurtado Salazar, A.; Pereira da Silva, D. F.; Nunes da Silveira Neto, A. 2023. Correlations between physical and chemical characteristics of Cortibel guava (*Psidium guajava* L.) fruits grown in the Brazilian Cerrado. *Revista de la Facultad de Ciencias Agrarias. Universidad Nacional de Cuyo. Mendoza. Argentina*. 55(1): 10-16. DOI: <https://doi.org/10.48162/rev.39.091>
46. Rombolà, A. D.; Quartieri, M.; Rodríguez-Declat, A.; Minnocci, A.; Sebastiani, L.; Sorrenti, G. 2023. Canopy-applied silicon is an effective strategy for reducing sweet cherry cracking. *Horticulture, Environment, and Biotechnology*. 64: 371-378. <https://doi.org/10.1007/s13580-022-00486-8>
47. Saba, M. K.; Sogvar, O.B. 2016. Combination of carboxymethyl cellulose-based coatings with calcium and ascorbic acid impacts in browning and quality of fresh-cut apples. *LWT-Food Science and Technology*. 66: 165-171. <https://doi.org/10.1016/j.lwt.2015.10.022>
48. Safa M.; Hajilou J.; Nagshiband-Hasani R.; Ganbari-Najar M. 2015. Effect of postharvest oxalic acid and calcium chloride on quality attributes of sweet cherry (*Prunus avium* L.). *Journal of Horticulture Science*. 29(2): 196-206. <https://doi.org/10.22067/JHORTS4.V010.29791>
49. Sharma, M.; Jacob, J. K.; Subramanian, J.; Paliyath, G. 2010. Hexanal and 1-MCP treatments for enhancing the shelf life and quality of sweet cherry (*Prunus avium* L.). *Scientia Horticulturae*. 125(3): 239-247. <https://doi.org/10.1016/j.scienta.2010.03.020>
50. SIAP. Sistema de Información Agrícola y Pesquera. 2023. <https://nube.siap.gob.mx/cierreagricola/> (Accesed August 2023).
51. Sohail, M.; Ayub, M.; Khalil, S. A.; Zeb, A.; Ullah, F.; Afridi, S. R.; Ullah, R. 2015. Effect of calcium chloride treatment on postharvest quality of peach fruit during cold storage. *International Food Research Journal*. 22(6): 2225-2229.
52. Toivonen, P. M. A. 2014. Relationship of typical core temperatures after hydrocooling on retention of different quality components in sweet cherry. *HortTechnology*. 24: 457-462. <https://doi.org/10.21273/HORTTECH.24.4.457>
53. Toivonen, P. M. A.; Manganaris, G. A. 2020. Chapter 15.2 - Stone fruits: Sweet cherries (*Prunus avium* L.), Editor(s): Maria Isabel Gil, Randolph Beaudry. *Controlled and modified atmospheres for fresh and fresh-cut produce*. Academic Press. 323-328. <https://doi.org/10.1016/B978-0-12-804599-2.00018-1>
54. Tsaniklidis, G.; Kafkaletou, M.; Delis, C.; Tsantili, E. 2017. The effect of postharvest storage temperature on sweet cherry (*Prunus avium* L.) phenolic metabolism and colour development. *Scientia Horticulturae*. 225: 751-756. <https://doi.org/10.1016/j.scienta.2017.08.017>

55. Tsantili, E.; Rouskas, D.; Christopoulos, M. V.; Stanidis, V.; Akrivos, J.; Papanikolaou, D. 2007. Effects of two pre-harvest calcium treatments on physiological and quality parameters in 'Vogue' cherries during storage. *The Journal of Horticultural Science and Biotechnology*. 82(4): 657-663. <https://doi.org/10.1080/14620316.2007.11512287>
56. Valero, D.; Pérez-Vicente A.; Martínez-Romero D.; Castillo S.; Guillén F.; Serrano M. 2002. Plum storability improved after calcium and heat postharvest treatments: Role of polyamines. *Journal of Food Science*. 67(7): 2571-2575. <https://doi.org/10.1111/j.1365-2621.2002.tb08778.x>
57. Vangdal, E.; Hovland, K. L.; Børve, J.; Sekse, L.; Slimestad, R. 2006. Foliar application of calcium reduces postharvest decay in sweet cherry fruit by various mechanisms. *Acta Horticulturae*. 768: 143-148. <https://doi.org/10.17660/ActaHortic.2008.768.16>
58. Wang, Y.; Xie, X.; Long, L. E. 2014. The effect of postharvest calcium application in hydro-cooling water on tissue calcium content, biochemical changes, and quality attributes of sweet cherry fruit. *Food Chemistry*. 160: 22-30. <https://doi.org/10.1016/j.foodchem.2014.03.073>
59. Wang, Y.; Long, L. E. 2015a. Physiological and biochemical changes relating to postharvest splitting of sweet cherries affected by calcium application in hydrocooling water. *Food Chemistry*. 181: 241-247. <https://doi.org/10.1016/j.foodchem.2015.02.100>
60. Wang, Y.; Bai, J.; Long, L. E. 2015b. Quality and physiological responses of two late-season sweet cherry cultivars 'Lapins' and 'Skeena' to modified atmosphere packaging (MAP) during simulated long distance ocean shipping. *Postharvest Biology and Technology*. 110: 1-8. <https://doi.org/10.1016/j.postharvbio.2015.07.009>
61. Wani, A. A.; Singh, P.; Gul, K.; Wani, M. H.; Langowski, H. C. 2014. Sweet cherry (*Prunus avium*): Critical factors affecting the composition and shelf life. *Food Packaging Shelf*. 1: 86-99. <https://doi.org/10.1016/j.fpsl.2014.01.005>
62. Wei, J.; Qi, X.; Guan, J.; Zhu, X. 2011. Effect of cold storage and 1-MCP treatment on postharvest changes of fruit quality and cell wall metabolism in sweet cherry. *Journal of Food Agriculture and Environment*. 9: 118-122. <https://doi.org/10.1234/4.2011.2235>
63. Winkler, A.; Knoche, M. 2019. Calcium and the physiology of sweet cherries: A review. *Scientia Horticulturae*. 245: 107-115. <https://doi.org/10.1016/j.scienta.2018.10.012>
64. Winkler, A.; Knoche, M. 2021. Calcium uptake through skins of sweet cherry fruit: effects of different calcium salts and surfactants. *Scientia Horticulturae*. 276: 109761. <https://doi.org/10.1016/j.scienta.2020.109761>
65. Winkler, A.; Fiedler, B.; Knoche, M. 2020. Calcium physiology of sweet cherry fruits. *Trees*. 34: 1157-1167. <https://doi.org/10.1007/s00468-020-01986-9>
66. Zhao, H.; Wang, B.; Cui, K.; Cao, J.; Jiang, W. 2019. Improving postharvest quality and antioxidant capacity of sweet cherry fruit by storage at near-freezing temperature. *Scientia Horticulturae*. 246: 68-78. <https://doi.org/10.1016/j.scienta.2018.10.054>