

# Seed treatments with salicylic acid and *Azospirillum brasiliense* enhance growth and yield of maize plants (*Zea mays L.*) under field conditions

## Tratamientos de semillas con ácido salicílico y *Azospirillum brasiliense* aumentan el crecimiento y el rendimiento de las plantas de maíz (*Zea mays L.*) en condiciones de campo

Luis Alfredo Rodríguez Laramendi <sup>1\*</sup>, Miguel Ángel Salas-Marina <sup>1</sup>,  
Vidal Hernández García <sup>1</sup>, Rady Alejandra Campos Saldaña <sup>1</sup>, Wel Cruz Macías <sup>1</sup>,  
Raúl López Sánchez <sup>2</sup>

Originales: Recepción: 24/08/2022 - Aceptación: 27/02/2023

### ABSTRACT

Salicylic acid and *Azospirillum brasiliense* stimulate plant growth and productivity. In some environments, plant physiology similarly responds to both bioactive products. Considering this, a field experiment was conducted to study the physiological effect of Salicilic acid and *A. brasiliense* on growth and grain yield of maize plants. The experiment involved three treatments consisting of imbibed seeds in an aqueous solution of SA (0.01 mM), inoculated seeds with *A. brasiliense* and a control treatment. Seed imbibition in SA and inoculation with *A. brasiliense* improved vegetative growth in the early stages of crop ontogeny, increasing leaf growth, plant height, stem diameter and biomass accumulation. Spikelet length and weight were greater in plants first inoculated with *A. brasiliense* and then treated with SA. Results indicated that SA stimulated biomass partitioning towards leaves, root and stem, while *A. brasiliense* mainly affected leaf growth, plant height, ear dimensions and grain yield. Such results turn crucial for biological fertilization strategies aimed at reducing pollutant loads that accompany chemical fertilizers. Both products can be part of maize management practices given competitive economic advantages and sustainability.

### Keywords

bioactive products • biofertilizers • plant hormones

1 Universidad de Ciencias y Artes de Chiapas. Facultad de Ingeniería Campus Villa Corzo. Carretera a Monterrey Km 3.0. Villa Corzo CP 30520. Chiapas. México.

\* alfredo.rodriguez@unicach.mx

2 Universidad de Granma. Carretera a Peralejo Km 18. Bayamo 85100. Granma. Cuba.

## RESUMEN

El ácido salicílico y *Azospirillum brasiliense* estimulan el crecimiento y la productividad de las plantas y, en algunos entornos, las respuestas fisiológicas de las plantas a la aplicación de ambos productos bioactivos parecen ser similares. Con el objetivo de estudiar su efecto fisiológico en el crecimiento y la producción de granos de plantas de maíz se diseñó un experimento de campo. Los tratamientos consistieron en embeber las semillas en una solución acuosa de SA (0,01 mM) e inocularlas con *A. brasiliense* y un tratamiento control. La imbibición de las semillas en SA y la inoculación con *A. brasiliense* mejoraron el crecimiento vegetativo en las primeras etapas de la ontogenia del cultivo, aumentando el crecimiento foliar, la altura de la planta, el diámetro del tallo y la acumulación de biomasa. La longitud y el peso de las espigas fueron mayores en las plantas inoculadas con *A. brasiliense* y tratadas con SA, en ese orden. Los resultados indicaron que el SA estimuló la acumulación de biomasa hacia las hojas, la raíz y el tallo, mientras que *A. brasiliense* tuvo más influencia en el crecimiento de las hojas, la altura de la planta, las dimensiones de la mazorca y el rendimiento del grano. Tales resultados son relevantes para el diseño de estrategias de fertilización biológica con la respectiva reducción de la carga contaminante que acompaña a los fertilizantes químicos. Ambos productos pueden formar parte de las prácticas agronómicas del cultivo de maíz por sus ventajas económicas competitivas y por ser ambientalmente sostenibles.

### **Palabras clave**

productos bioactivos • biofertilizantes • hormonas vegetales

## **INTRODUCTION**

Maize constitutes an essential nutrient for the human diet. Despite a low protein content compared to other vegetables, it provides, on average, 39% of overall protein intake, and 59% of human energy requirements (38).

Mexico is the largest market for maize, consuming 11% of world production (29). However, the low-yielding domestic production does not meet the growing demand, and importation reaches approximately 11.3 and 0.145 million tons of yellow and white maize, respectively (36). Hence, there is an evident need to explore technological alternatives for environmentally friendly production and increasing yields.

The indiscriminate use of ammonia fertilizers on some bedrock types of soil, causes serious acidification problems. Soils in the Frailesca region of Chiapas are undergoing an acidification process (11) and consequent increasing toxic aluminium contents in maize plants, particularly at pH levels below 5.0 (6). Faced with such a situation, finding alternatives entailing reduced agrochemical applications may contribute to reduced soil degradation. In this sense, several biofertilizers based on nitrogen-fixing microorganisms have shown positive effects on certain cultivated plants, such as maize (3, 25, 32), sorghum (19), soybeans (10), and rice (12).

Among nitrogen-fixing bacteria, *A. brasiliense* is considered one of the most important plant growth-promoting bacteria (8). On one hand, this bacterium promotes growth and increases plant production, after promoting phytohormone synthesis (5) and biological nitrogen fixation. On the other hand, *A. brasiliense* restricts certain plant pathogens through antibiosis and siderophores (14). In addition, *Azospirillum* may associate with more than 100 plant species, 14 of which are grasses (30). As stated by Gavilanes *et al.* (2020), *A. brasiliense* (Ab-V5 and Ab-V6) has been widely used as a commercial inoculant in Brazil (14), with positive effects on grain dry matter and nitrogen accumulation in plants, especially in grain crops such as maize and wheat (13, 24).

Given the mentioned phytohormone-producing capacity of this bacterium, inoculation effects could be related to the role played by salicylic acid. Several studies performed with rhizosphere bacteria show that most strains produced metabolites of the AIA type, siderophores and salicylic acid (22). Salicylic acid (SA) is a phenolic secondary metabolite (20) present in plant tissues (33, 41), regulating plant growth and increasing crop yield when supplied in low exogenous concentrations (34). In maize, the application of SA increases grain production per plant, total dry biomass, and N, P, K contents (41).

Some hypotheses about the effect of SA and *A. brasiliense* consider the role of SA as a plant hormone and the ability of *A. brasiliense* to fix atmospheric nitrogen and increase nutrient uptake by promoting increased stem growth and grain production (15). The effect of both bioactive products (SA and *A. brasiliense*), effective biomolecules and plant growth regulators (26), depended on the genotype and environmental conditions. In this sense, given the agronomic implications on possible biological fertilization strategies in maize, comparing the effect of both products turns interesting. Thus, this research aimed to study the physiological effect of *A. brasiliense* and SA on growth and grain production of maize plants under field conditions.

## MATERIALS AND METHODS

### Location

The research was conducted from June 2016 to January 2017 in the locality of Calzada Larga, municipality of Villaflor, Chiapas, México, at 16°21'08.5" N and 93°18'58.2" W and 713 m a. s. l. (8). The predominant climates are warm and semi-warm, with an average temperature of 24.5 °C, an average rainfall of 1200 mm per year, and the following predominant soil types: lithosols, luvisols, cambisols and vertisols.

### Growth conditions and plant material

Seeds of the CLTHW11002 hybrid, yellow maize from the International Maize and Wheat Improvement Center (CIMMYT), were manually sowed at 0.8 m between rows and 0.35 m between plants on a luvisol soil with loamy texture and enough water content.

According to the Mexican Official Standard (27), the soil is strongly acidic, free of carbonates and salts, low in potassium, moderately low in organic matter, medium phosphorus (P-Bray) and nitrogen (N-NO<sub>3</sub>), moderately high in magnesium (Mg) and very high in iron (Fe). Both aluminium content and proportion vs. total soil cations (4.61%) were low, despite the extreme soil acidity (table 1).

**Table 1.** Soil chemical properties.

**Tabla 1.** Caracterización química del suelo del área experimental.

O. M: organic matter,  
CEC: cation exchange capacity.  
M. O: Materia Orgánica,  
CEC: Capacidad de intercambio catiónico.

<b>pH</b>	<b>O.M (g kg<sup>-1</sup>)</b>	N-NO <sub>3</sub>	P-Bray	K	Ca	Mg	Fe	Al	<b>CEC (Meq/100g)</b>
		<b>mg kg<sup>-1</sup></b>							
4.49	14.4	16.6	25.4	1.03	2402	655	48.4	95.2	23.0

Two nitrogen fertilizations were applied at 15 days and 30 days after sowing, both at a rate of 75 kg ha<sup>-1</sup> of N, for a total dose of 150 kg ha<sup>-1</sup> (46-00-00). Weed chemical control was carried out with Velquat 1.5 L ha<sup>-1</sup> at 15 days after sowing, and 1.5 L ha<sup>-1</sup> Tacsquat at 60 days after sowing. The fall armyworm (*Spodoptera frugiperda*) was controlled with Cipermetrine 21.12% EC at a dose of 1.0 L ha<sup>-1</sup>.

### Experimental design and treatments

The randomized block experimental design with three treatments and three replicates consisted of T1: salicylic acid (SA), T2: *A. brasiliense* (5 x 108 CFU), T3: control treatment without SA and not inoculated. Each experimental unit measured 25 m<sup>2</sup> (5 x 5 m), leaving one border plant on each side of the plot.

### **Seed inoculation with *A. brasiliense***

The commercial product tested was Azofer®, composed of *A. brasiliense* (50%), peat (37.5%) and calcium carbonate (12.5%). Calcium carbonate allows achieving the ideal pH (6.8-7.0) for the inoculant (6), while not affecting plant growth or development. The concentration of *A. brasiliense* was 5 x 108 CFU's per gram of commercial product.

For seed inoculation, an inert solution was prepared with 60 g of carbosil-methyl-cellulose adhesive powder dissolved in 1.5 L of distilled water and left to stand for two hours. Subsequently, moistened seeds with the adherent powder were homogeneously covered with the biofertilizer and left to dry in the shade, at room temperature, before sowing.

### **Imbibition of seeds with salicylic acid**

A solution with distilled water and salicylic acid at a concentration of 0.01 mM, was prepared according to Hayat and Ahmad (2007), Gordillo-Curiel *et al.* (2020) and Rodríguez-Laramendi *et al.* (2017). The seeds were soaked for 2 hours and left to dry at room temperature before sowing.

### **Analytical evaluations**

For growth measurements at 30 and 60 days after sowing, five plants per replicate were selected, number of leaves per plant (LP) was counted, and plant leaf area was determined in cm<sup>2</sup> (LA) with a CI-202 (Bioscience ®) portable leaf area meter. Plant height (PH) was measured with a millimetre ruler. Leaves, stem, roots of the five selected plants were separated, and oven dried at 80°C for 72 hours. Finally, leaves dry weight (LDW), shoot dry weight (SDW) and root dry weight (RDW) were obtained using a Sartorius® analytical balance. Plant dry weight was calculated.

### **Growth rates**

Based on leaf area and dry weight, the leaf area index (LAI) was calculated by dividing plant leaf area (LA) by soil coverage. Root weight fraction (RWF, g g<sup>-1</sup>) was calculated as the root dry weight vs. plant dry weight ratio. This indicator expresses the "root's investment" or biomass gain (4). Leaf weight fraction (LWF, g g<sup>-1</sup>) was calculated as leaf dry weight vs. plant dry weight, estimating leaf partitioning (42). The root mass/leaf mass ratio (dimensionless) estimated phenotypic plasticity in biomass allocation (4). Specific leaf area (SLA, cm<sup>2</sup> g<sup>-1</sup>) was calculated as leaf area/leaf dry weight, reflecting functional traits of leaf morphology such as thickness and density (31) and vegetative vigor (23). Leaf area ratio (LAR, cm<sup>2</sup> g<sup>-1</sup>), one important determinant of plant relative growth rate (42), was calculated as leaf area by plant dry weight.

### **Yield components**

When the crop reached physiological maturity, five plants per treatment were selected in each replicate and ear length (EL, cm), ear diameter (ED, cm), 100-grain dry weight, and dry grain yield per plant (Yield, g plant<sup>-1</sup>) at 14% grain moisture, were assessed.

### **Statistical analysis**

ANOVA with a randomized block design followed by LSD mean comparison test was conducted considering p ≤ 0.05. ANOVA assumptions were verified by the Cochran and Bartlett tests. A multivariate Principal Component Analysis (PCA) searched for a possible relationship between plant growth, yield, ear weight and ear size. All analyses were performed with the statistical package STATISTICA® release 8.0 (39).

## **RESULTS**

The number of leaves per plant was significantly higher in inoculated plants, compared to those treated with SA and control, the latter of which produced fewer leaves. Despite a tendency towards more leaves per plant on both sampling dates (at 30 and 60 das), at 60 das, this difference was not significant (table 2, page 21).

**Table 2.** Effect of salicylic acid (SA) and *A. brasiliense* on growth of maize plants at 30 and 60 days after sowing (das).**Tabla 2.** Efecto del ácido salicílico (AS) y *A. brasiliense* en el crecimiento de plantas de maíz a los 30 y 60 días después de la siembra (dds).

Treatments	Leaves per plant (LP)	Plant height (PH, cm)	Stem diameter (SD, cm)	Leaf area (LA, m <sup>2</sup> plant <sup>-1</sup> )	Leaf dry weight (LDW, g)	Stem dry weight (SDW, g)	Root dry weight (RDW, g)	Plant dry weight (PDW, g)
30 das								
SA	8.75 b	48.04 a	1.96 a	0.14 a	5.01 a	3.11	2.12 a	10.24 a
<i>A. brasiliense</i>	9.42 a	44.29 b	1.77 b	0.12 ab	5.02 a	3.26	1.67 ab	9.96 ab
Control	8.00 c	42.93 b	1.58 c	0.10 b	3.96 b	2.86	1.40 b	8.21 b
Standard error	0.11	0.50	0.04	0.01	0.23	0.25	0.15	0.52
60 das								
SA	10.08	168.00 a	1.98 a	0.37	33.04 a	56.83 a	37.88 a	127.75 a
<i>A. brasiliense</i>	10.17	165.00 a	1.80 b	0.34	29.42 b	53.63 b	32.21 b	115.25 b
Control	9.00	155.83 b	1.63 c	0.32	24.96 c	43.21 c	25.92 c	94.08 c
Standard error	0.36	2.17	0.04	0.02	0.83	2.02	1.96	3.55

ns: Not significant; \* Statistically significant for  $p \leq 0.05$ , das: days after sowing

Ns: sin diferencias significativas,  
\* Diferencias estadísticas significativas para  $p \leq 0.05$ , dds: días después de la siembra.

However, plant height and stem diameter were significantly higher in plants grown from SA-treated seed, and even higher than those inoculated with *A. brasiliense* at 30 das. At 60 das, no significant difference in plant height was observed between SA and *A. brasiliense*-treated plants (table 2).

Leaf area was larger in SA-treated plants at 30 das than control plants. The latter exhibited no significant differences concerning those inoculated with *A. brasiliense*. At 60 das, no difference in leaf area was observed among treatments (table 2).

Leaf dry weight was statistically higher in plants treated with SA and *A. brasiliense* than in control plants at 30 das. At 60 das this difference was even higher for SA-treated plants followed by *A. brasiliense*, compared to the control. Similar results were observed for root and stem dry weight and, therefore, for total plant dry weight. However, stem dry weight was statistically different at 60 das (table 2).

Table 3 (page 22) shows the effect of bioactive products on growth ratios, only significant at 30 das, when an increasing leaf area index was observed for the SA treatment. No difference was observed between *A. brasiliense* and control. At 60 das, such growth-stimulating effects were not significant (table 3, page 22).

The higher leaf growth values observed with SA and *A. brasiliense* at early stages of maize plant development (table 2) was similar to the increase detected in ear length and weight, and yield per plant (figure 1, page 22).

**Table 3.** Effect of the application of salicylic acid (SA) and *A. brasiliense* on growth rates of maize plants at 30 and 60 days after sowing (das).**Tabla 3.** Efecto de la aplicación de ácido salicílico (AS) y *A. brasiliense* en los índices de crecimiento de plantas de maíz a los 30 y 60 días después de la siembra (dds).

Treatments	Leaf area index (LAI)	Specific leaf area ( $\text{cm}^2 \text{g}^{-1}$ )	Leaf area ratio ( $\text{g g}^{-1}$ )	Leaf weight ratio ( $\text{g g}^{-1}$ )	Leaf weight fraction ( $\text{g g}^{-1}$ )	Root/stem ratio ( $\text{g g}^{-1}$ )	Root/leaf weight ratio ( $\text{g g}^{-1}$ )
30 das							
SA	0.50 a	285.16	138.19	0.21 a	0.49	0.27 a	0.43
<i>A. brasiliense</i>	0.44 ab	248.21	124.89	0.17 b	0.50	0.20 b	0.34
Control	0.37 b	262.77	127.58	0.17 b	0.48	0.21 b	0.36
Standard error	0.03	14.46	8.53	0.01	0.01	0.02	0.03
60 das							
SA	1.31	112.24	29.11	0.29	0.26	0.42	1.14
<i>A. brasiliense</i>	1.23	116.73	29.77	0.28	0.25	0.39	1.10
Control	1.15	130.03	34.68	0.27	0.27	0.38	1.04
Standard error	0.06	5.22	1.99	0.01	0.01	0.02	0.04

Ns: Not significant;

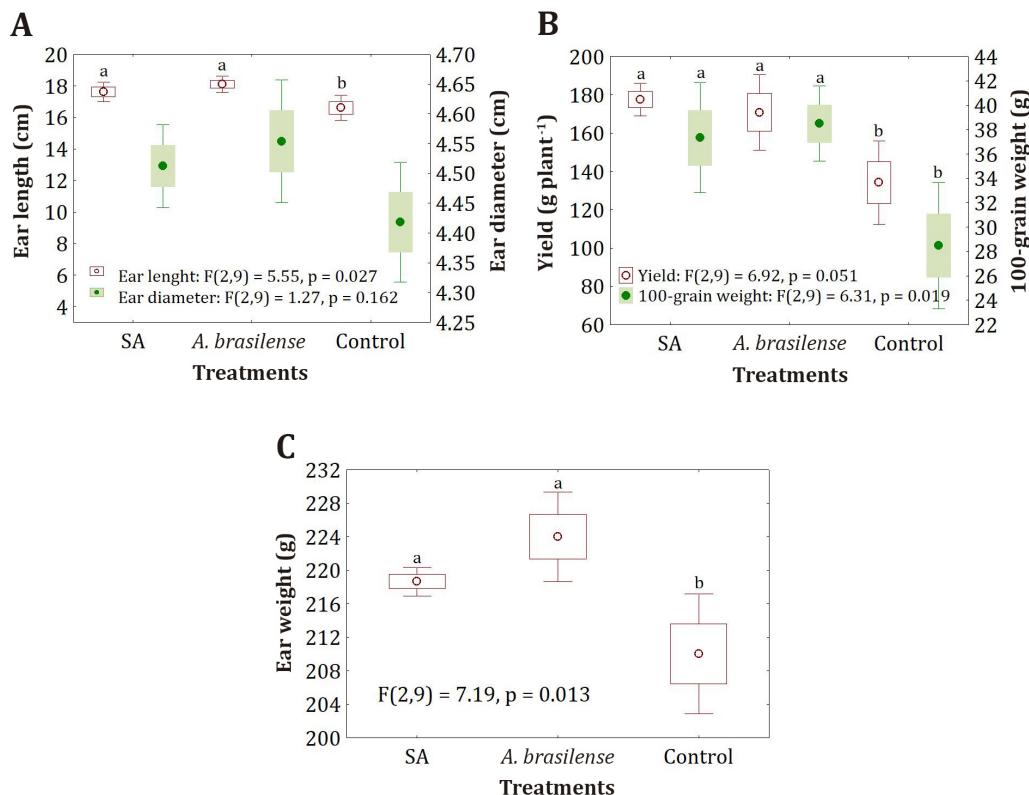
\* Statistically significant for  $p \leq 0.05$ .

Ns: sin diferencias significativas,

\* Diferencias estadísticas

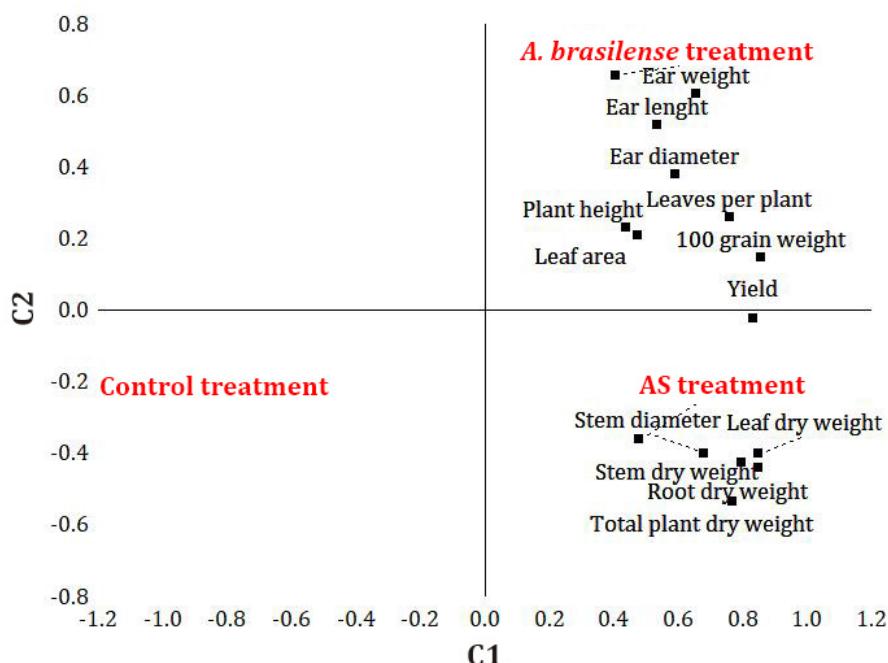
significativas para  $p \leq 0.05$  dds: días

después de la siembra.

**Figure 1.** Effect of salicylic acid and *A. brasiliense* on A) ear length and diameter, B) yield and 100-grain weight and C) ear weight.**Figura 1.** Efecto del tratamiento con ácido salicílico y *A. brasiliense* en: A) longitud y diámetro de la mazorca, B) rendimiento y peso de 100 granos y C) peso de la mazorca.

Plants treated with SA and *A. brasiliense* developed longer ears, consistently with the higher 100-grain weight (figure 1 B, page 22), ear weight and yield per plant (figure 1 B, C, page 22).

Considering vegetative growth, ear size and grain production, a PCA allowed establishing the effects of both bioactive products as a function of the evaluated variables (figure 2). Component 1 (48.58%) discriminates control plants from both bioactive treatments, and component 2 (16.20%) discriminates both bioactive treatments. Variables related to biomass accumulation and stem diameter, discriminate in favour of SA treatment, while variables related to leaf growth, plant height, ear dimensions and grain yield, separate the *A. brasiliense* treatment (figure 2).



**Figure 2.** PCA biplot of growth variables, ear dimensions and grain yield of corn plants treated with SA and *A. brasiliense*, according to components C1 (48.58%) and C2 (16.20%).

**Figura 2.** Biplot de las variables de crecimiento, dimensiones de la mazorca y producción de granos de plantas de maíz tratadas con AS y *A. brasiliense*, en el plano formado por los componentes C1 (48.58%) y C2 (16.20%).

## DISCUSSION

Maize growth response to seed inoculation with *A. brasiliense* and imbibition with SA on two sampling dates suggests a possible relationship with plant physiological stage and ontogeny, something to be further addressed in future research. The effect of both bioactive products resulted organ dependent.

The results further demonstrate that SA and *A. brasiliense* have similar effects, promoting vegetative growth of maize plants at the early stages. However, these effects depend on sampling date (34) and particularly, on plant organ. This effect is probably given by SA and *A. brasiliense* sharing an analogous property, stimulating hormone synthesis and interacting with cytokinins. SA constitutes a plant hormone exceeding plant immunity and abiotic stress (34). In coordination with cytokinins, ethylene, auxins, gibberellins, jasmonic acid and abscisic acid, SA significantly contributes to growth and development regulation, although through unknown mechanisms (34). In this sense, it has been shown that the

growth-promoting effects of SA could be related to hormone modulation (1, 37) or gas exchange improvement (39).

The PCA showed correlations between leaf growth *vs.* plant height, and ear size *vs.* increased grain production. These aspects are related to the *A. brasiliense* enhanced nitrogen absorption, even in fertile soils (15), and enhanced hormone synthesis, including auxin, gibberellin, and cytokinin (2, 17).

The effect of SA on grain production in maize plants is also supported by other studies. Low doses of SA induce increased ear length and maize grain yield (41). However, in our research, SA was applied to leaves (41) and not seeds, opening a new field of research.

Positive plant responses to inoculation with *A. brasiliense* might be given by biological nitrogen fixation, probably overshadowed by the nitrogen fertilization carried out in the experiment, and by plant hormonal production. In this regard, inoculations with *A. brasiliense* had improved root growth and development of *Setaria viridis* grass after increased CO<sub>2</sub> fixation and reduced accumulation of photo-assimilated carbon in leaves, resulting in greater canopy growth, increased water content in plant tissues, and reduced stress (24, 28). In addition, increased production of indoleacetic acid may improve nutrient uptake by augmenting root growth (24, 28). Zeffa *et al.* (2019) studied *A. brasiliense* inoculated seeds, finding intensified plant growth, improved biochemical traits and raised NUE under nitrogen deficit. Other authors have found that regardless of nitrogen source and dosage, *A. brasiliense* increased maize grain yield (16).

SA effects on plant dry weight are probably related to SA ability to increase N, P<sub>2</sub>O<sub>5</sub> and K<sub>2</sub>O contents in plant tissues (41). On the other hand, seed inoculation with *A. brasiliense* stimulates root growth, increases root exploration capacity and promotes biological nitrogen fixation, which may be related to higher grain production and ear length in inoculated plants. Specifically, in maize, other studies have shown that inoculations with nitrogen-fixing bacteria result in a 9% increase in maize grain production (16).

Our results show that SA and *A. brasiliense* had similar stimulating effects on growth and yield of maize plants, from early stages of plant ontogeny. This physiological effect may be related to the ability to interact with other hormones (37) or promote hormone synthesis (16). However, we suggest caution before issuing a hypothesis on this matter as, apart from the above-mentioned properties, *A. brasiliense* contributes to plant nutrition through biological nitrogen fixation, a fact that may mask the effects of both bioactive products on plant growth and development.

## CONCLUSIONS

We consider that the results obtained show robust evidence supporting theoretical and methodological bases, as well as sufficient evidence on the physiological effect of SA and the bacterium *A. brasiliense* on growth and grain production of maize under field conditions. Such results are relevant to the design of biological fertilization strategies with the respective reduction of pollutants. Considering economic and environmental sustainability, both products can be considered within the agronomic practices of maize cultivation.

The results indicated that SA and *A. brasiliense* exhibited similar stimulating effects on growth and yield of maize plants, from early stages. SA induced greater plant biomass accumulation, with a tendency to maximize root dry mass, while *A. brasiliense* stimulated greater leaf growth and plant height, consequently inducing increased ear growth and grain production.

## REFERENCES

1. Abreu, M. E.; Munné-Bosch, S. 2009. Salicylic acid deficiency in NahG transgenic lines and sid2 mutants increases seed yield in the annual plant *Arabidopsis thaliana*, Journal of Experimental Botany. 60: 1261-1271.
2. Bashan, Y.; de-Bashan, L. E. 2010. How the plant growth-promoting bacterium *Azospirillum* promotes plant growth - a critical assessment. Adv. Agron. 108: 77-136. doi: 10.1016/S0065-2113(10)08002-8
3. Bertasello, L. E. T.; Filla, V. A.; Prates Coelho, A.; Vitti Môro, G. 2021. Agronomic performance of maize (*Zea mays*) genotypes under *Azospirillum brasiliense* application and mineral fertilization. Revista de la Facultad de Ciencias Agrarias. Universidad Nacional de Cuyo. Mendoza. Argentina. 53(1): 68-78.
4. Camargo, I.; Rodríguez, N. 2006. New perspectives to study the biomass allocation and its relationship with the functioning of plants in neotropical ecosystems. Acta biológica colombiana. 11: 75-87.
5. Carcaño-Montiel, M. G.; Ferrera-Cerrato, R.; Pérez-Moreno, J.; Molina-Galán, D.; Bashan, Y. 2016. Actividad nitrogenasa, producción de fitohormonas, sideróforos y antibiosis en cepas de *Azospirillum* y *Klebsiella* aisladas de maíz y teocintle. Terra Latinoamericana. 24: 493-502.
6. Casierra, F.; Aguilar, O. Estrés por aluminio en las plantas: reacciones en el suelo, síntomas en vegetales y posibilidades de corrección. 2007. Revista Colombiana de Ciencias Hortícolas. 1: 246-256.
7. Cassán, F.; Penna, C.; Creus, C.; Radovancich, D.; Monteleone, E.; Salamone, I. G.; Lett, L. 2010. Protocolo para el control de calidad de inoculantes que contienen *Azospirillum* sp. Buenos Aires, Argentina: Asociación Argentina de Microbiología. [consultado el 1 de febrero de 2023].
8. Cassán, F. D.; Díaz-Zorita, M. 2016. *Azospirillum* sp. in current agriculture: from the laboratory to the field. Soil Biol. Biochem. 103:117-130. doi: 10.1016/j.soilbio.2016.08.020
9. CEIEG. 2011. Ubicación Geográfica; Climas; Vegetación y Uso del Suelo; Edafología; Geología; Fisiografía; Hidrografía; Áreas Naturales Protegidas. Población. Villaflóres, Chiapas.
10. Chibeba, A. M.; Guimarães, M. D. F.; Brito, O. R.; Nogueira, M. A.; Araujo, R. S.; Hungria, M. 2015. Co-inoculation of soybean with *Bradyrhizobium* and *Azospirillum* promotes early nodulation. Am. J. Plant Sci. 6:1641-1649. <https://doi.org/10.4236/ajps.2015.610164>
11. Cruz-Macías, O. W.; Rodríguez-Laramendi, R. L.; Salas-Marina, M. A.; Hernández-García, V.; Campos-Saldaña, R. A.; Chávez-Hernández, M. H.; Gordillo-Curiel, A. 2020. Efecto de la materia orgánica y la capacidad de intercambio catiónico en la acidez de suelos cultivados con maíz en dos regiones de Chiapas, México. Terra Latinoamericana. 38: 475-480. DOI: <https://doi.org/10.28940/terra.v38i3.506>
12. El-Khawas, H.; Adachi, K. 1999. Identification and quantification of auxins in culture media of *Azospirillum* and *Klebsiella* and their effect on rice roots. Biol. Fertil. Soils. 28: 377-381.
13. Fukami, J.; Nogueira, M. A.; Araujo, R. S.; Hungria, M. 2016. Accessing inoculation methods 327 of maize and wheat with *Azospirillum brasiliense*. AMB Express. 6(1): 3.
14. Fukami, J.; Abrantes, J. L. F.; del Cerro, P.; Nogueira, M. A.; Ollero, F. J.; Megías, M.; Hungria, M. 2017. Revealing strategies of quorum sensing in *Azospirillum brasiliense* strains 325 Ab-V5 and Ab-V6. Archives of Microbiology: 1-10. 326
15. Galindo, F. S.; Teixeira-Filho, M. C. M.; Buzetti, S.; Santini, J. M. K.; Alves, J. M. K.; Ludkiewicz, M. G. Z. 2017. Wheat yield in the Cerrado as affected by nitrogen fertilization and inoculation with *Azospirillum brasiliense*. Pesqui. Agropecu. Bras. 52: 794-805. doi:10.1590/s0100-204x2017000900012
16. Galindo, F. S.; Teixeira Filho, M. C. M.; Buzetti, S.; Pagliari, P. H.; Santini, J. M. K. 2019. Maize yield response to nitrogen rates and sources associated with *Azospirillum brasiliense*. Agron J. 111: 1985-1997. <https://doi.org/10.2134/agronj2018.07.0481>
17. Gavilanes, F. Z.; Souza Andrade, D.; Zucareli, C.; Horácio, E. H.; Sarkis Yunes, J.; Barbosa, A. P.; Ribeiro Alves, L. A.; García Cuzatty, L.; Raju Maddela, N.; Guimarães, M. de F. 2020. Co-inoculation of *Anabaena cylindrica* with *Azospirillum brasiliense* increases grain yield of maize hybrids. Rhizosphere. 15: 100224. doi: 10.1016/j.rhisph.2020.100224
18. Gordillo Curiel, A.; Rodríguez Laramendi, L. A.; Salas Marina, M. Á.; Rosales Esquinca, M. de los Á. 2020. Efecto del ácido salicílico sobre la germinación y crecimiento inicial del café (*Coffea arabica* var. Costa Rica 95). Revista de la Facultad de Agronomía de la Universidad del Zulia. 38(1): 43-59. Retrieved from <https://produccioncientificafaluz.org/index.php/agronomia/article/view/34729>
19. Grellet-Naval, N.; Vera, L.; Leggio, N. F.; Fernández, G. P.; Sánchez, D. A.; Fernández U. J.; Romero, E. R.; Tórtora, M. L. 2017. Evaluación de la cepa *Azospirillum brasiliense* Az39 como biofertilizante para el cultivo de sorgo azucarado. Revista Industrial y Agrícola de Tucumán. 94: 31-39.
20. Hayat, Q.; Hayat, S.; Irfan, M.; Ahmad, A. 2010. Effect of exogenous salicylic acid under changing environment: a review. Environ. Exp. Bot. 68: 14-25. doi: 10.1016/j.envexpbot.2009.08.005
21. Hayat, S.; Ahmad, A. 2007. Salicylic Acid a Plant Hormone. Springer Publishers, Dordrecht, The Netherlands.
22. Hernández, A.; Rives, N.; Caballero, A.; Hernández, A. N.; Heydrich, M. 2004. Caracterización de rizobacterias asociados al cultivo del maíz en la producción de metabolitos del tipo AIA, sideróforos y ácido salicílico, Rev. Col. Biotec. 6: 6-13.

23. Hugalde, I.; Riaz, S.; Agüero, C. B.; Romero, N.; Barrios-Masias, F.; Nguyen, A. V.; Vila, H.; McElrone, A.; Gómez Talquena, S.; Arancibia, C.; Walker, M. A. 2017. Physiological and genetic control of vigour in a 'Ramsey' × 'Riparia Gloire de Montpellier' population. *Acta Horticulturae*. 1188: 205-212.
24. Hungria, M.; Campo, R. J.; Souza, E. M.; Pedrosa, O. F. 2010. Inoculation with selected strains of *Azospirillum brasiliense* and *A. lipoférum* improves yields of maize and wheat in Brazil. *Plant Soil*. 331: 413-425. <https://doi.org/10.1007/s11104-009-0262-0>
25. Montejo-Martínez, D.; Casanova-Lugo, F.; García-Gómez, M.; Oros-Ortega, I.; Díaz-Echeverría, V.; Morales-Maldonado, E. R. 2018. Foliar and radical response of maize to biological-chemical fertilization in a Luvisol soil. *Agronomía Mesoamericana*. 29: 325-341. <https://dx.doi.org/10.15517/ma.v29i2.29511>
26. Nandy, S.; Das, T.; Dey, A. 2021. Role of jasmonic acid and salicylic acid signaling in secondary metabolite production. In: Aftab, T.; Yusuf, M. (eds). *Jasmonates and salicylates signaling in plants. signaling and communication in plants*. Springer. Cham. [https://doi.org/10.1007/978-3-030-75805-9\\_5](https://doi.org/10.1007/978-3-030-75805-9_5)
27. Norma Oficial Mexicana NOM-021-RECNAT-2000. 2002. Norma Oficial Mexicana que establece las especificaciones de fertilidad, salinidad y clasificación de suelos. Estudios, muestreos y análisis. Diario Oficial de la Federación. México, D. F.
28. Pankievicz, V. C. S.; do Amaral, F. P.; Santos, K. F. D. N.; Agtuca, B.; Xu, Y.; Schueler, M. J.; Arisi, A. C. M.; Steffens, M. B.; de Souza, E. M.; Pedrosa, F. O.; Stacey, G.; Ferrieri, R. A. 2015. Robust biological nitrogen fixation in a model grass–bacterial association. *Plant J.* 81: 907-919. doi:10.1111/tpj.12777
29. Peña-Datoli, M.; Hidalgo-Moreno, C.; González-Hernández, V.; Alcántar-González, E.; Etchevers-Barra, J. 2016. Recubrimiento de semillas de maíz (*Zea mays L.*) con quitosano y alginato de sodio y su efecto en el desarrollo radical. *Agrociencia*. 50: 1091-1106.
30. Pereg, L.; de-Bashan, L. E.; Bashan, Y. 2016. Assessment of affinity and specificity of *Azospirillum* for plants. *Plant Soil*. 399: 389-414. <https://doi.org/10.1007/s11104-015-2778-9>
31. Poorter, H.; Nagel, O. 2000. The role of biomass allocation in the growth response of plants to different levels of light, CO<sub>2</sub>, nutrients and water: a quantitative review. *Australian Journal of Plant Physiology*. 27(12): 595-607. doi: 10.1071/PP99173\_CO
32. Rangel-Lucio, J. A.; Rodríguez-Mendoza, M. N.; Ferrera-Cerrato, R.; Castellanos-Ramos, J. Z.; Ramírez-Gama, R. M.; Alvarado-Bárcenas, E. 2011. Afinidad y efecto de *Azospirillum* spp. en maíz. *Agronomía Mesoamericana*. 22(2): 269-279.
33. Raskin, I. 1992. Role of salicylic acid in plants. *Ann. Rev. Plant Physiol. Molec. Biol.* 43: 439-463.
34. Rivas-San Vicente, M.; Plasencia, J. 2011. Salicylic acid beyond: its role in plant growth and development. *J. Exp. Bot.* 62: 1-18.
35. Rodríguez-Larramendi, L. A.; Ramírez, M. G.; Gómez-Rincón, M. A.; Guevara-Hernández, F.; Salas-Marina, M. Á.; Gordillo-Curiel, A. 2017. Efectos del ácido salicílico en la germinación y crecimiento inicial de plántulas de frijol (*Phaseolus vulgaris L.*). *Rev. Fac. Agron. (LUZ)*. 34: 253-269.
36. Servicio de Información Agroalimentaria y Pesquera (SIAP). 2021. Producción agrícola. Gobierno de México. <https://www.gob.mx/siap/acciones-y-programas/produccion-agricola-33119> (fecha de consulta: 02/08/2022).
37. Shakirova, F. M.; Sakhabutdinova, A. R.; Bezrukova, V.; Fatkhutdinova, R. A.; Fatkhutdinova, D. R. 2003. Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. *Plant Science*. 164: 317-322.
38. Sierra-Macías, M.; Becerra-Leor, E. N.; Palafax-Caballero, A.; Barrón-Freyre, S.; Cano-Reyes, O.; Zambada-Martínez, A.; Sandoval-Rincón, A.; Romero-Mora, J. 2004. Caracterización de híbridos de maíz (*Zea mays L.*) con alta calidad de proteína por su rendimiento y tolerancia a pudrición de mazorca en el sureste de México. *Revista Mexicana de Fitopatología*. 22: 268-276.
39. StatSoft, Inc. 2007. STATISTICA (data analysis software system), version 8.0. [www.statsoft.com](http://www.statsoft.com).
40. Stevens, J.; Senaratna, T.; Sivasithamparam, K. 2006. Salicylic acid induces salinity tolerance in tomato (*Lycopersicon esculentum* cv. Roma): Associated changes in gas exchange, water relations and membrane stabilisation. *Plant Growth Regul.* 49: 77-83. <https://doi.org/10.1007/s10725-006-0019-1>
41. Tucuch-Hass, C.; Alcántara, G. G.; Trejo, T. L. I.; Volke, H. H.; Salinas, M. Y. y Larqué, S. A. 2017. Efecto del ácido salicílico en el crecimiento, estatus nutrimental y rendimiento en maíz (*Zea mays*). *Agrociencia*. 51: 771-781.
42. Villar, R.; Ruiz-Robleto, J.; Quero, J.; Poorter, H.; Valladares, F.; Marañón, T. 2004. Tasas de crecimiento en especies leñosas: aspectos funcionales e implicaciones ecológicas. En: Valladares, F. (Ed.), *Ecología del bosque mediterráneo en un mundo cambiante*. Madrid, España: Ministerio de Medio Ambiente. EGRAF, S. A. 193-230.
43. Zeffa, D. M.; Perini, L. J.; Silva, M. B.; de Sousa, N. V.; Scapim, C. A.; Oliveira, A. L. M. d.; Texeira do Amaral, J. A.; Simoes, A. G. L. 2019. *Azospirillum brasiliense* promotes increases in growth and nitrogen use efficiency of maize genotypes. *PLoS ONE*. 14(4): e0215332. <https://doi.org/10.1371/journal.pone.0215332>